

# Today's Lecture

- Failure of equal frequency assumption
- Neutralist vs selectionist interpretations
- Site models
- Comparing models: Likelihood ratios & weight matrices

# Failure of Equal Frequency Assumption for (Real) DNA

- For most organisms, the nucleotide composition is significantly different from .25 for each nucleotide, e.g.:
  - *H. influenza* .31 A, .19 C, .19 G, .31 T
  - *P. aeruginosa* .17 A, .33 C, .33 G, .17 T
  - *M. janaschii* .34 A, .16 C, .16 G, .34 T
  - *S. cerevisiae* .31 A, .19 C, .19 G, .31 T
  - *C. elegans* .32 A, .18 C, .18 G, .32 T
  - *H. sapiens* .29 A, .21 C, .21 G, .29 T

- Note approximate symmetry:  $A \cong T$ ,  $C \cong G$ ,
  - even though we're counting nucs on just one strand.
  - Expect *exact* equality when counting both strands
- Explanation:
  - Although individual biological features may have non-symmetric composition (local *asymmetry*),
  - usually features are distributed approx *randomly* w.r.t. strand,
  - so local asymmetries *cancel*, yielding overall symmetry.

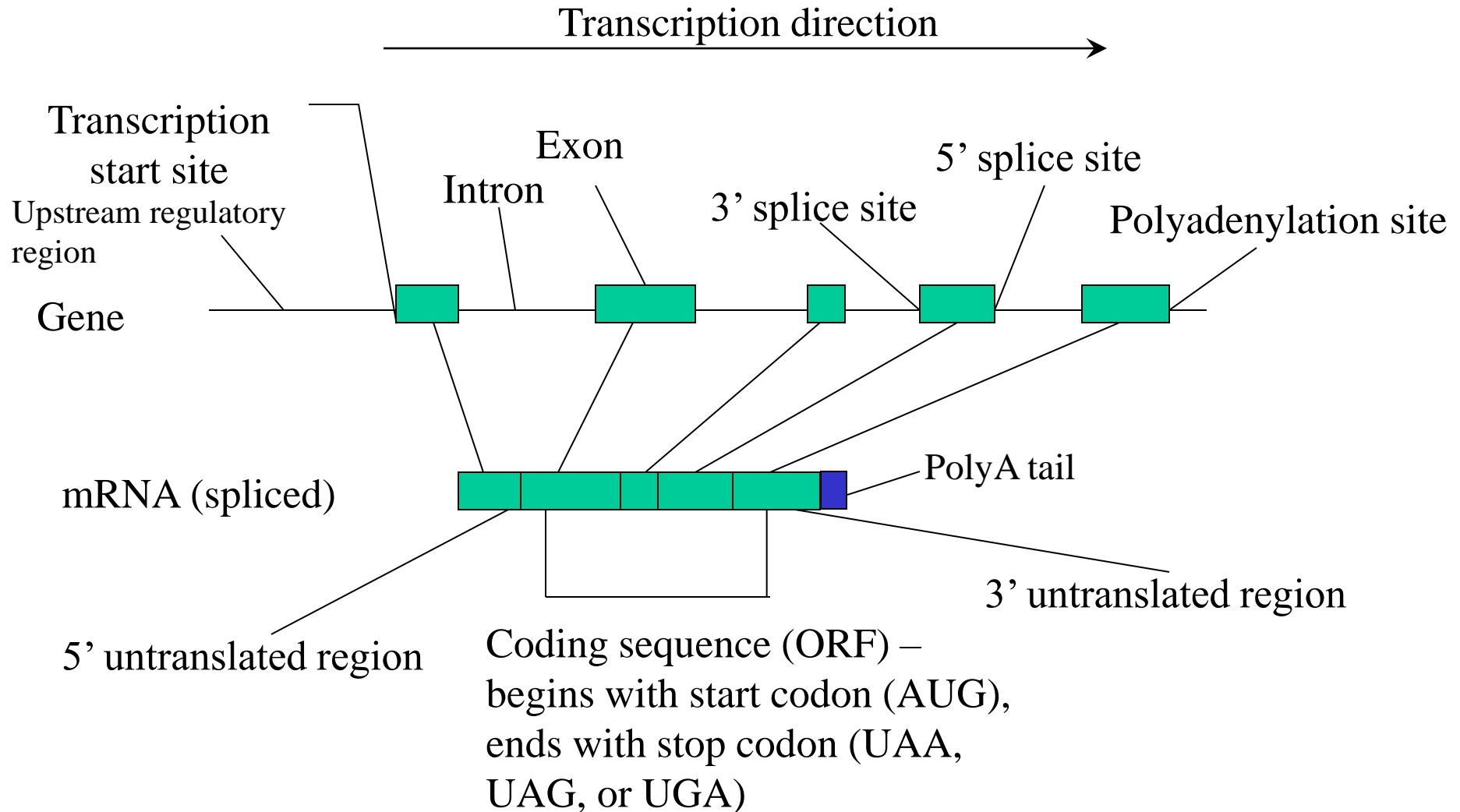
# General Hypotheses Regarding Unequal Frequency

- **Neutralist** hypothesis: *mutation bias*
  - e.g. due to nucleotide pool composition
- **Selectionist** hypothesis: *selection*
  - selection on (many) particular nucleotides
  - selection on mutational bias mechanisms
  - ...

# Site Models

- Probability models for short sequences, such as:
  - splice sites
  - translation start sites
  - promoter elements
  - protein “motifs”

# (Protein-coding) Gene Structure in Eukaryotes



- Assumptions:
  - different examples of site can be aligned *without gaps* (indels) such that tend to have same residues in same positions
  - drop equal freq assumption: allow *position-specific freqs*
  - retain *independence* assumption (for now)

- Applies to short segments (< 30 residues) where
  - precise residue spacing is structurally or functionally important, and
  - certain positions are highly conserved
- Examples:
  - DNA/RNA sequences binding a single protein or RNA molecule
  - Protein internal regions structurally constrained due to folding requirements; or
  - protein surface regions constrained because bind certain ligands

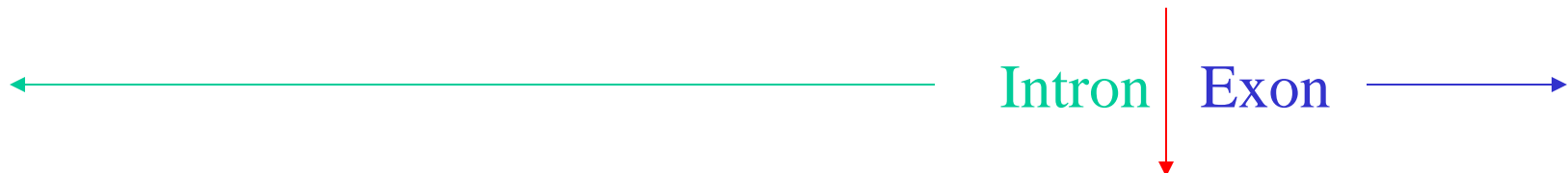


# Construction of Site Models

- Collect examples of site
- Align (without gaps)
- Count occurrences of residues at each position
- Convert to frequencies

# Nucleotide Counts for 8192 *C. elegans* 3' Splice Sites

3' ss



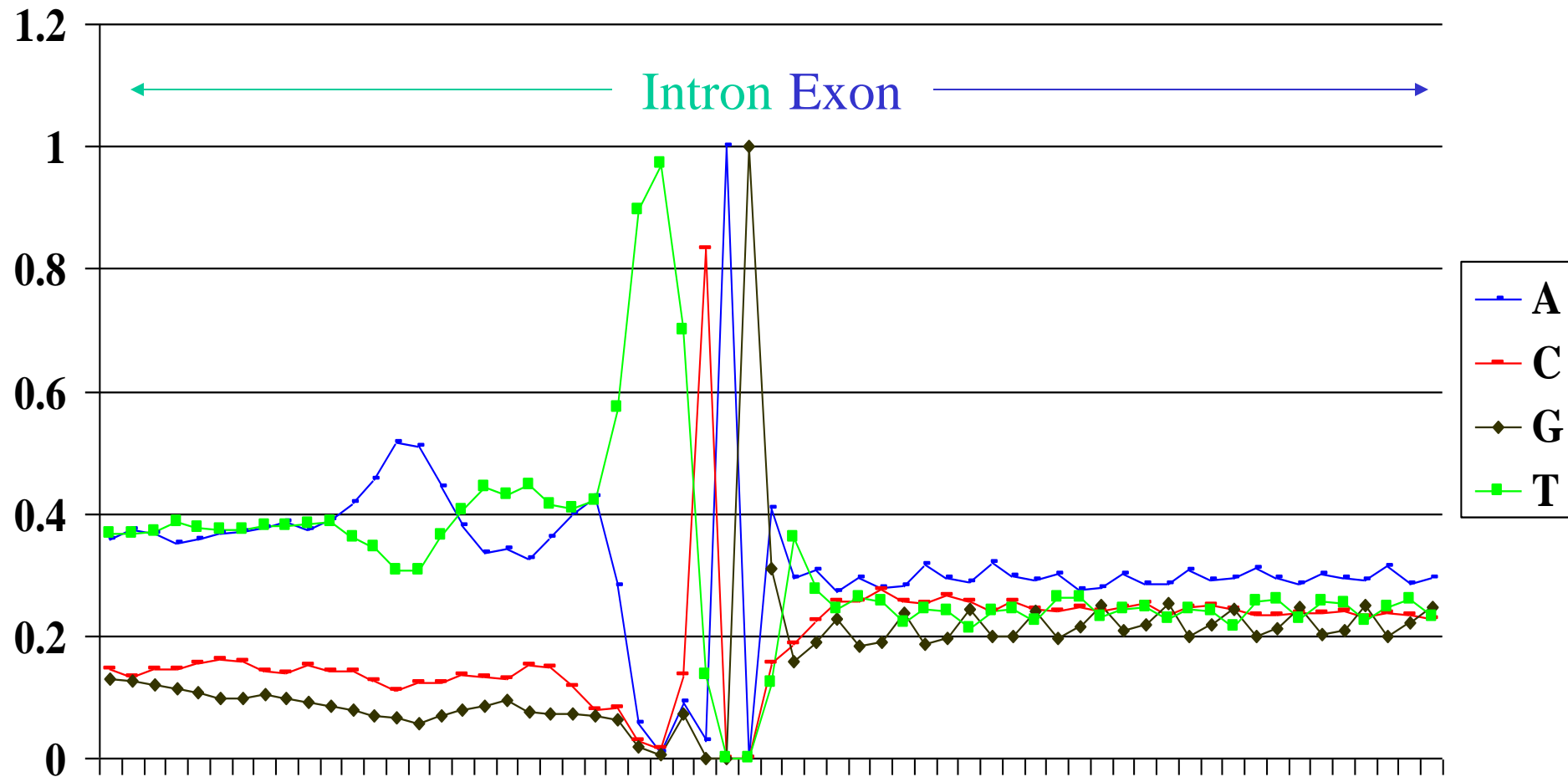
A	3276	3516	2313	476	67	757	240	8192	0	3359	2401	2514
C	970	648	664	236	129	1109	6830	0	0	1277	1533	1847
G	593	575	516	144	39	595	12	0	8192	2539	1301	1567
T	3353	3453	4699	7336	7957	5731	1110	0	0	1017	2957	2264

**CONSENSUS**

W W W T T t C A G r w w

A	0.400	0.429	0.282	0.058	0.008	0.092	0.029	1.000	0.000	0.410	0.293	0.307
C	0.118	0.079	0.081	0.029	0.016	0.135	0.834	0.000	0.000	0.156	0.187	0.225
G	0.072	0.070	0.063	0.018	0.005	0.073	0.001	0.000	1.000	0.310	0.159	0.191
T	0.409	0.422	0.574	0.896	0.971	0.700	0.135	0.000	0.000	0.124	0.361	0.276

# 3' Splice Sites – *C. elegans*



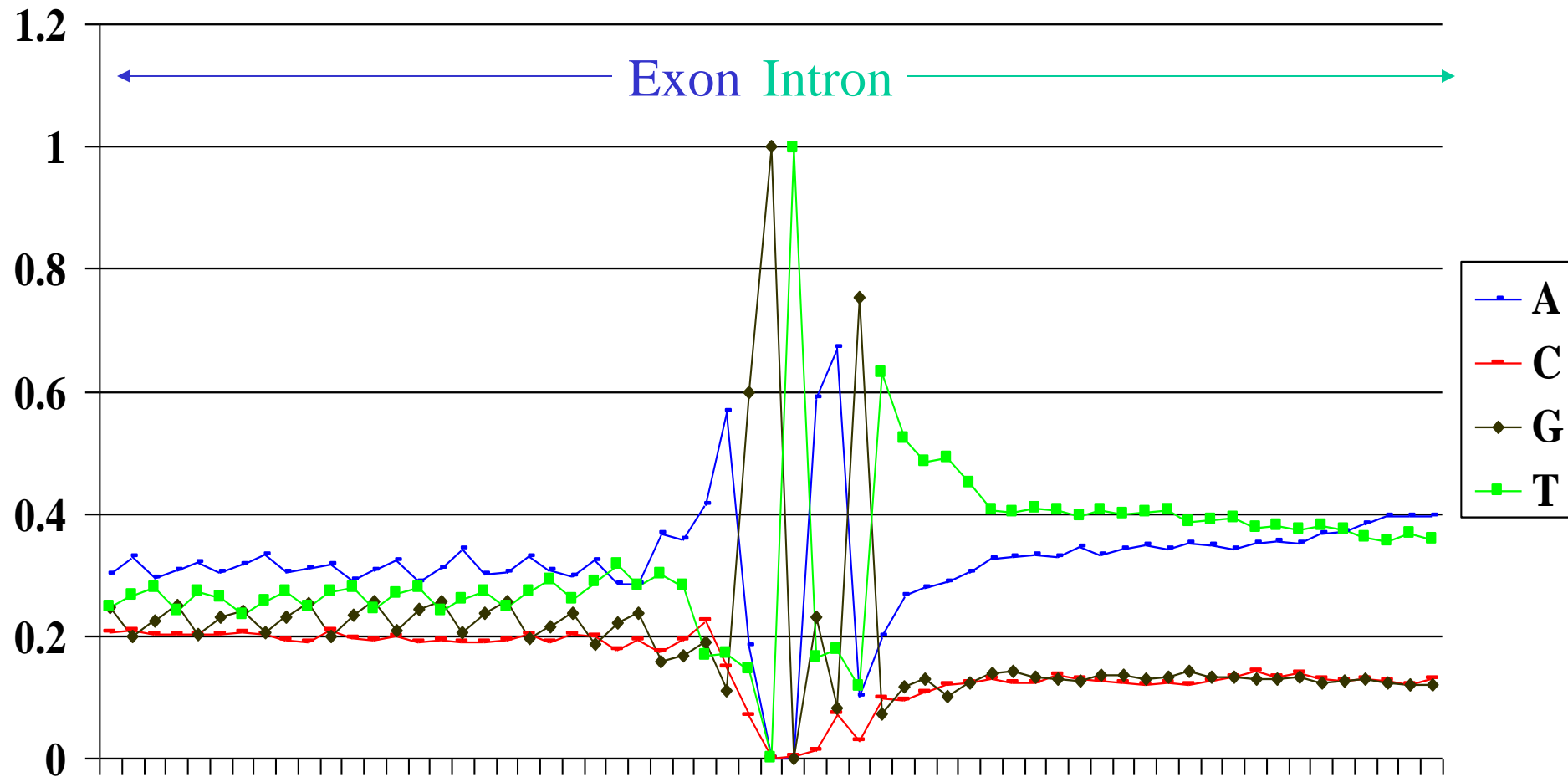
# Nucleotide Counts for 8192 *C. elegans* 5' Splice Sites



A	3404	4644	1518	0	0	4836	5486	837	1632	2189	2278	2355
C	1850	1224	583	0	14	118	588	237	801	771	889	986
G	1562	912	4891	8192	0	1890	672	6164	589	962	1056	827
T	1376	1412	1200	0	8178	1348	1446	954	5170	4270	3969	4024

CONSENSUS	x	a	g	G	T	a	a	g	t	t	w	t
A	0.416	0.567	0.185	0.000	0.000	0.590	0.670	0.102	0.199	0.267	0.278	0.287
C	0.226	0.149	0.071	0.000	0.002	0.014	0.072	0.029	0.098	0.094	0.109	0.120
G	0.191	0.111	0.597	1.000	0.000	0.231	0.082	0.752	0.072	0.117	0.129	0.101
T	0.168	0.172	0.146	0.000	0.998	0.165	0.177	0.116	0.631	0.521	0.484	0.491

# 5' Splice Sites – *C. elegans*



# Conserved Domain in RecR and Class I Topoisomerases

RecR RLAE EKITEVILATNPTEGEATANYIAELC  
 RecM RLQDDQVTEVILATNPNIERGEATAMYISRLL  
 RecR RVDDVGITEVILATDPNTEGEATATYLVVMV  
 TrsI IFKENKIDEVILATDPAREGENIAYKILNQL  
 TOP1 KQLAEKADHIYLATDL DREG EAI AWRLREVI  
 ORF1 AELLKQANTIIVATDSDREGENIAWSIIHKA  
 TOP1 KDALKDADELILATDEDREGKVISWHLLQLL  
 TOP1 TIFDKRVKTIILATDAAAEGEYIGRNILYRL  
 TOP3 KREARNADYLMIWTD CDREGEYIGWEIWQEA  
 TOP3 KRFLHEASEIVHAGDPDREGQLLVDEVLDYL  
 RGYR RNLAVEADEVLIGTDPDTEGEKIAWDLYLAL

**CONSENSUS**    **xxxxxxxxxxU&uatDxxxEGexxxxxUxxxu**

*Consensus key:*

**Uppercase:** all residues chemically similar

**lowercase:** most are

**U,u:** bulky aliphatic (I,L,V)

**&:** bulky hydrophobic (I,L,V,M,F,Y,W)

From RL Tatusov, SF Altschul, and EV Koonin, PNAS 91: 12091-12095

# Probability Models for Sites (assuming independence!)

- For each position  $i$ ,  $1 \leq i \leq n$ , let  $P_i$  be a prob dist'n on the alphabet of residues
  - e.g. constructed using counts at that position in a sample of sites.
  - $P_i(r)$  for each residue  $r$  is the probability that  $r$  occurs at position  $i$  in a sequence.
- Prob dist'n  $P$  on the space  $S$  of sequences of length  $n$  is defined by

$$P(s) = \prod_{1 \leq i \leq n} P_i(s_i)$$

where  $s = s_1 s_2 \dots s_n$

# Zero Probabilities

- If  $P_i(r) = 0$  for some  $i$  and  $r$ , then  $P(s) = 0$  for some sequences.
  - may or may not be desirable
- If due to failure to observe residue because of small sample size,
  - should perform “small-sample correction” to change  $P_i(r)$  to a small non-zero value.
  - usually done by adding ‘pseudocounts’ to each value in the counts matrix;
    - e.g. add 1 to each cell (has justification in Bayesian statistics)
  - Particularly an issue with proteins, due to larger alphabet size.
- If reflects real biological constraints
  - then leave as 0.
  - e.g. requirement for G at position +1 (first intronic base) in 5’ss



# Comparing Alternative Probability Models

- We will want to consider more than one model at a time, in following situations:
  - To differentiate between two or more hypotheses about a sequence
  - To generate increasingly refined probability models that are progressively more accurate

- First situation arises in testing biological assertion, e.g. “is this a coding sequence?”
  - Compare two models:
    1. model associated with a hypothesis  $H_{coding}$ ,
      - assigns each sequence the prob of observing it under expt of drawing a coding sequence at random from genome
    2. model associated with a hypothesis  $H_{noncoding}$ ,
      - assigns each sequence the prob of observing it under expt of drawing a non-coding sequence at random

# Likelihood Ratios

- The *likelihood* of a model  $M$  given an observation  $s$  is

$$L(M | s) = P(s | M)$$

This is *not* the *probability* of the model! – (the sum over all models is not 1).

- The *likelihood ratio* ( $LR$ ) of two models  $M_a$  and  $M_0$  is given by

$$LR(M_a, M_0 | s) = \frac{L(M_a | s)}{L(M_0 | s)}$$

The numerator and denominator may both be very small!

- The *log likelihood ratio* ( $LLR$ ) is the logarithm of the likelihood ratio.

# Weight Matrices for Site Models

- LR for sites: (prob under site model) / (prob under non-site (background) model)

$$\frac{P(s | M_{\text{site}})}{P(s | M_{\text{background}})} = \frac{\prod_{1 \leq i \leq n} P_i(s_i | M_{\text{site}})}{\prod_{1 \leq i \leq n} P_i(s_i | M_{\text{background}})}$$

- $\text{LLR} = \sum_{1 \leq i \leq n} \log(P_i(s_i | M_{\text{site}})) - \log(P_i(s_i | M_{\text{background}}))$ 
  - compute by reading from a *matrix* whose  $i$ -th column contains values  $\log(P_i(r | M_{\text{site}})) - \log(P_i(r | M_{\text{background}}))$  for each residue  $r$  (with  $r$  labelling the rows).
    - We use  $\log_2$ .

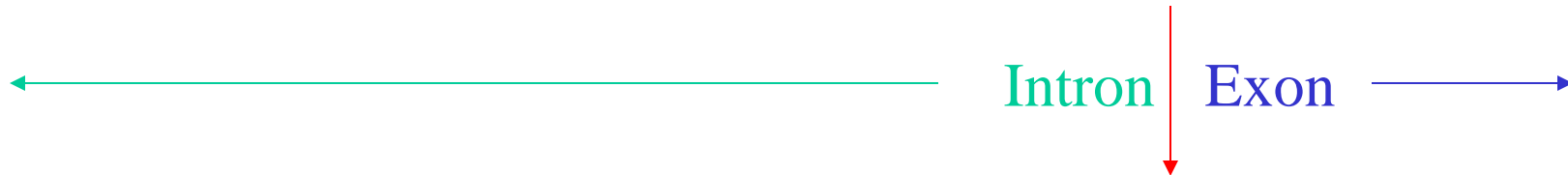
# Example: 3' splice sites in *C. elegans*

- For *background distribution* take
  - genomic residue freqs computed from *C. elegans* chrom. I:

A	4,575,132:	0.321
C	2,559,048:	0.179
G	2,555,862:	0.179
T	4,582,688:	0.321
  - other choices are possible, e.g. composition of *transcribed regions*
- For the *site distribution* we take
  - site residue freqs from 8192 sites:

# Nucleotide Counts for 8192 *C. elegans* 3' Splice Sites

3' ss



A	3276	3516	2313	476	67	757	240	8192	0	3359	2401	2514
C	970	648	664	236	129	1109	6830	0	0	1277	1533	1847
G	593	575	516	144	39	595	12	0	8192	2539	1301	1567
T	3353	3453	4699	7336	7957	5731	1110	0	0	1017	2957	2264

**CONSENSUS** W W W T T t C A G r w w

A	0.400	0.429	0.282	0.058	0.008	0.092	0.029	1.000	0.000	0.410	0.293	0.307
C	0.118	0.079	0.081	0.029	0.016	0.135	0.834	0.000	0.000	0.156	0.187	0.225
G	0.072	0.070	0.063	0.018	0.005	0.073	0.001	0.000	1.000	0.310	0.159	0.191
T	0.409	0.422	0.574	0.896	0.971	0.700	0.135	0.000	0.000	0.124	0.361	0.276

# Weight Matrix – 3' Splice Sites

## SITE FREQUENCIES:

A	0.400	0.429	0.282	0.058	0.008	0.092	0.029	1.000	0.000	0.410	0.293	0.307
C	0.118	0.079	0.081	0.029	0.016	0.135	0.834	0.000	0.000	0.156	0.187	0.225
G	0.072	0.070	0.063	0.018	0.005	0.073	0.001	0.000	1.000	0.310	0.159	0.191
T	0.409	0.422	0.574	0.896	0.971	0.700	0.135	0.000	0.000	0.124	0.361	0.276

## BACKGROUND FREQUENCIES:

A	0.321	0.321	0.321	0.321	0.321	0.321	0.321	0.321	0.321	0.321	0.321	0.321
C	0.179	0.179	0.179	0.179	0.179	0.179	0.179	0.179	0.179	0.179	0.179	0.179
G	0.179	0.179	0.179	0.179	0.179	0.179	0.179	0.179	0.179	0.179	0.179	0.179
T	0.321	0.321	0.321	0.321	0.321	0.321	0.321	0.321	0.321	0.321	0.321	0.321

## WEIGHTS:

A	0.32	0.42	-0.18	-2.46	-5.29	-1.79	-3.45	1.64	-99.00	0.36	-0.13	-0.06
C	-0.60	-1.18	-1.15	-2.64	-3.51	-0.41	2.22	-99.00	-99.00	-0.20	0.06	0.33
G	-1.31	-1.35	-1.51	-3.35	-5.23	-1.30	-6.93	-99.00	2.48	0.79	-0.17	0.10
T	0.35	0.39	0.84	1.48	1.60	1.12	-1.24	-99.00	-99.00	-1.37	0.17	-0.22

# Scoring a Candidate 3' Splice Site

A	0.32	0.42	-0.18	-2.46	-5.29	-1.79	-3.45	1.64	-99.00	0.36	-0.13	-0.06
C	-0.60	-1.18	-1.15	-2.64	-3.51	-0.41	2.22	-99.00	-99.00	-0.20	0.06	0.33
G	-1.31	-1.35	-1.51	-3.35	-5.23	-1.30	-6.93	-99.00	2.48	0.79	-0.17	0.10
T	0.35	0.39	0.84	1.48	1.60	1.12	-1.24	-99.00	-99.00	-1.37	0.17	-0.22

T T C T T A C A G A A T

$$0.35 + 0.39 + -1.15 + 1.48 + 1.60 + -1.79 + 2.22 + 1.64 + 2.48 + 0.36 + -0.13 + -0.22 = 7.23$$



- General def.: a *weight matrix*  $W$  has entries  $w_{rj}$  indexed by residues  $r \in A$ , and  $1 \leq j \leq n$
- *score* of a sequence  $s = (s_1 s_2 \dots s_n)$  is

$$\sum_{1 \leq j \leq n} w_{s_j j}$$

- In the site case,

$$w_{rj} = \log(P_j(r | M_{\text{site}})) - \log(P_j(r | M_{\text{background}}))$$