

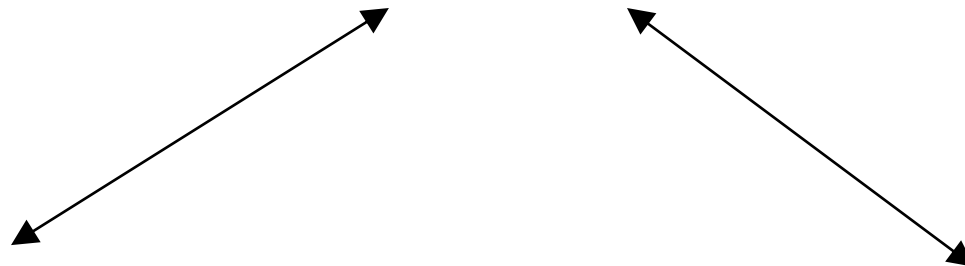
# Lecture 1.1: Overviews

- Computational molecular biology
- Genome biology
- Genomicists' tasks
- Role of computation
- This course

# Computational Molecular Biology

## Molecular biology

*poses questions, judges answers*



Probability and statistics

Computational *models* for  
biological processes

Computer science

*Methods* for computation:  
Computers & languages  
Data structures & algorithms

- This course is about *sequence-based* CMB
  - i.e. methods (& models) for obtaining & analyzing the information encoded in the genome
- We do not cover ‘non-linear’ (non-sequence based) computational biology, such as:
  - Most proteomics, metabolic & signalling pathways, models for interacting molecules ...

# Genome biology overview

- Genomes undergo two fundamental processes (both involve copying!):
  - Replication
  - Transcription
- Genomic functional information is in the form of *sites*:
  - Short (~2 – ~15 base) sequence segments that bind to an *RNA* or *protein* molecule (the *reader*) to help mediate some function

# Two broad classes of sites:

1. Sites acting *at the DNA level* (usually via *protein* readers) to help carry out or regulate a fundamental process
  - Replication
    - Replication origins, centromeres, telomeres (each usually having *multiple* sites)
  - Transcription
    - Promoters, enhancers, suppressors (each usually having *multiple* sites, with readers being *transcription factors*)

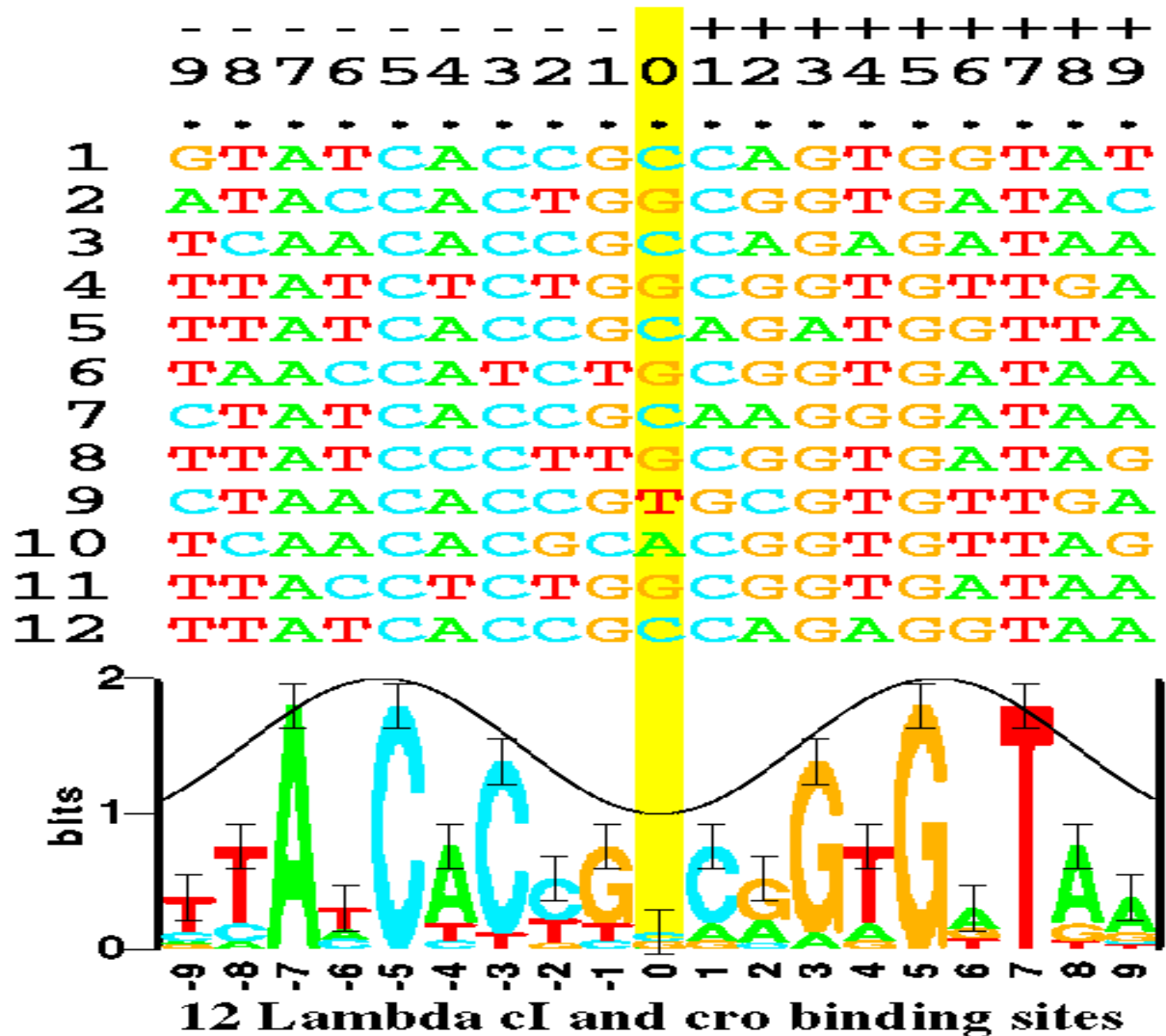
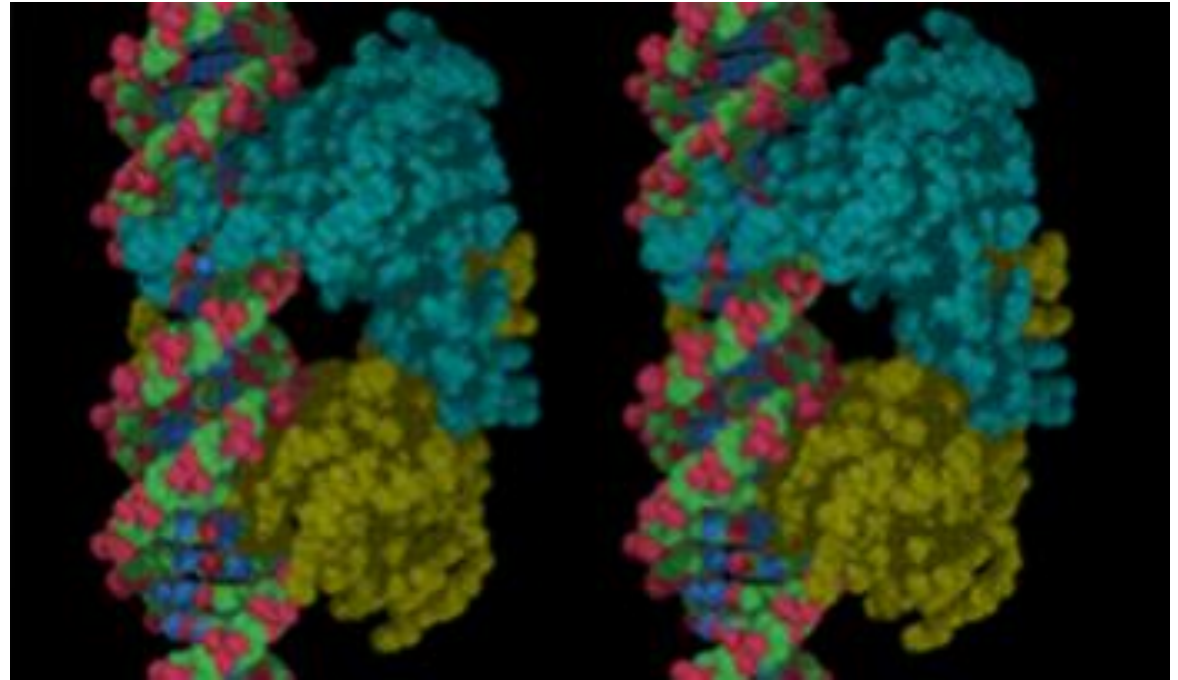
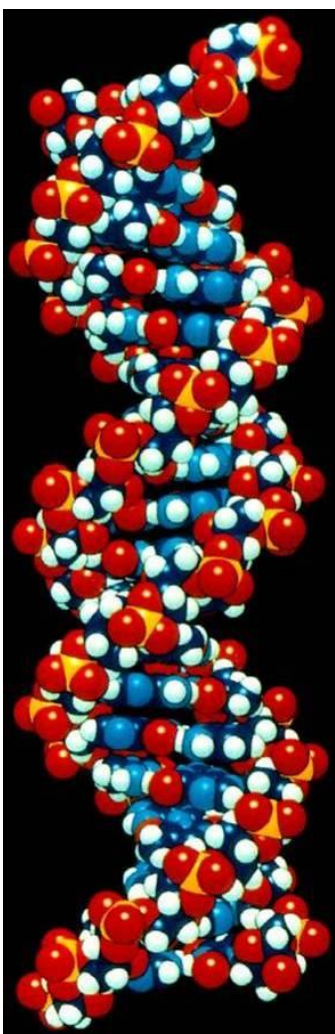


Fig. 1. Some aligned sequences and their sequence logo. At the top of the figure are listed the 12 DNA sequences from the  $P_L$  and  $P_R$  control regions in bacteriophage lambda. These are bound by both the  $cI$  and  $cro$  proteins [16]. Each even numbered sequence is the complement of the preceding odd numbered sequence. The sequence logo, described in detail in the text, is at the bottom of the figure. The cosine wave is positioned to indicate that a minor groove faces the center of each symmetrical protein. Data which support this assignment are given in reference [17].



*from* <http://gibk26.bse.kyutech.ac.jp>

*from* <http://www.dna-dna.net/>

2. Sites acting *within a transcript* (often via RNA readers) to help carry out the transcript's function

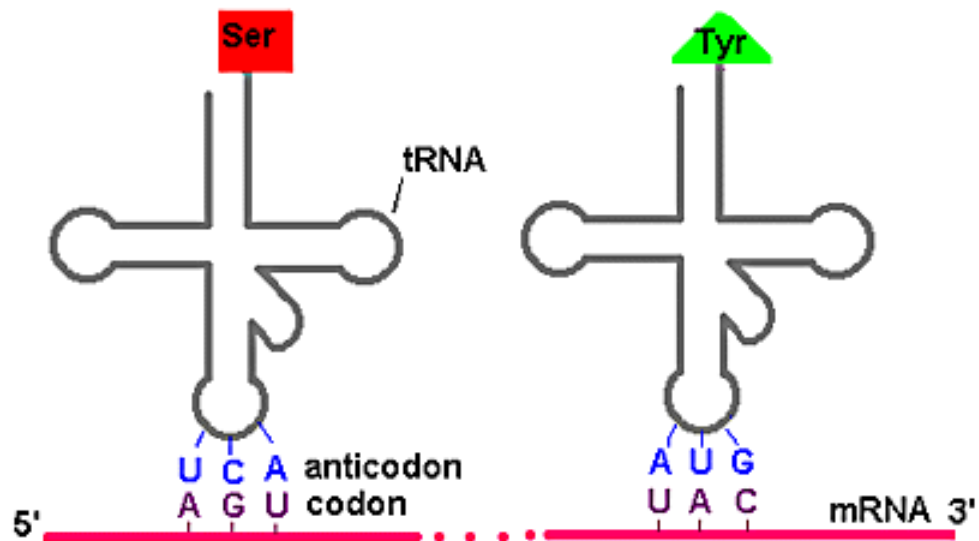
– in *protein coding* transcripts:

- Translation start sites, codons (reader = charged tRNA), splice sites, microRNA binding sites, polyadenylation sites, ...

– in *functional RNA* transcripts:

- Stem structures (the transcript reads itself!), ...



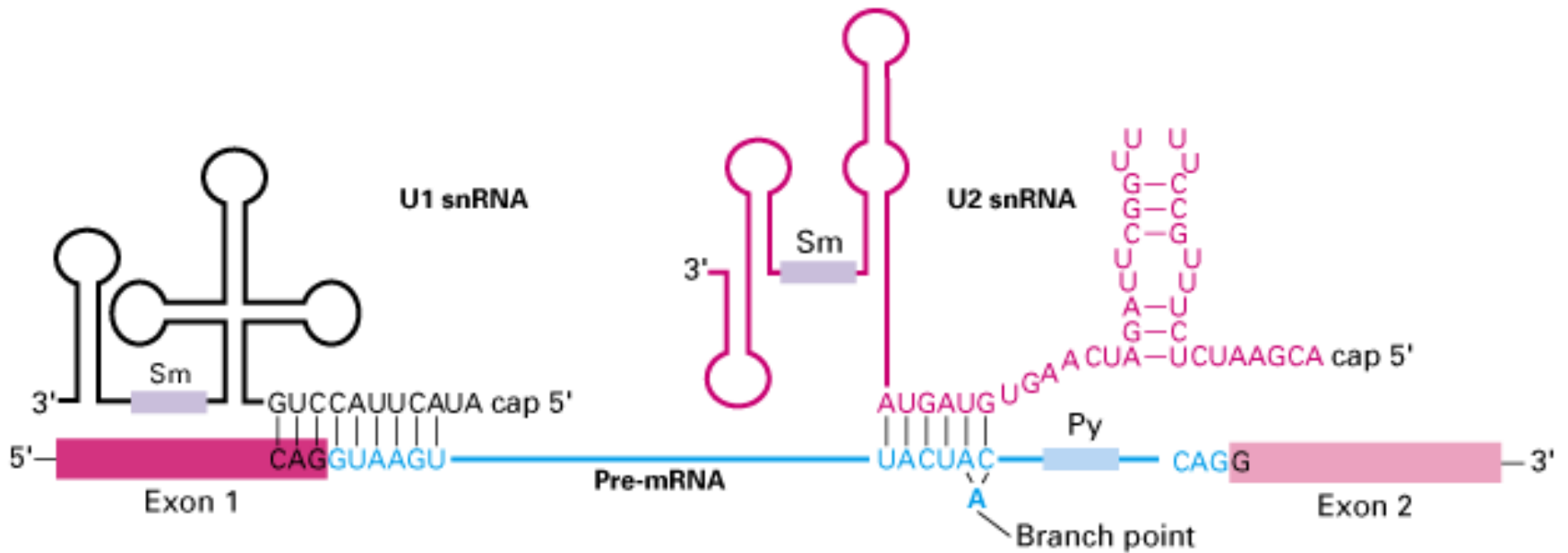


2nd base in codon

		U	C	A	G	
1st base in codon	U	Phe Phe Leu Leu	Ser Ser Ser Ser	Tyr Tyr STOP STOP	Cys Cys STOP Trp	U C A G
	C	Leu Leu Leu Leu	Pro Pro Pro Pro	His His Gln Gln	Arg Arg Arg Arg	U C A G
	A	Ile Ile Ile Met	Thr Thr Thr Thr	Asn Asn Lys Lys	Ser Ser Arg Arg	U C A G
	G	Val Val Val Val	Ala Ala Ala Ala	Asp Asp Glu Glu	Gly Gly Gly Gly	U C A G

3rd base in codon

## The Genetic Code



from [http://departments.oxy.edu/biology/Stillman/bi221/111300/processing\\_of\\_hnrnas.htm](http://departments.oxy.edu/biology/Stillman/bi221/111300/processing_of_hnrnas.htm)

(Jonathon Stillman, Grace Fisher-Adams )

# Sites: some key properties

- Sites typically
  - *Recur*: multiple sites within a genome, with possibly varying sequences, may be recognized by the same reader
    - Sequence variation may be represented by a *motif* or *sequence logo*
  - *Cluster*: several sites, with the same or different readers, may act collectively to carry out some function
    - Positional constraints within clusters vary in stringency
    - A *gene* is a cluster of sites involved in *expressing* a particular transcript

- Average site density (i.e. the fraction of the genome that is functional) may be quite small!
  - $< 10\%$  of human genome; remaining  $> 90\%$  mostly transposon relics, dead genes & processed pseudogenes
- Whether or not a site is active in a given cell may depend on
  - reader status (local concentration, whether modified, etc)
    - Also interaction partners of the reader
  - chromatin & methylation status
  - whether nearby (or overlapping) sites are bound by their readers

# Genomicists' tasks

- Find the *genome sequence*
- Find the *transcripts*
- Find the *sites* ...
- ... and their *functions* ...

# Finding the genome sequence

- Get *reads* (short, overlapping, error-prone pieces of the sequence)
- *Assemble* : identify read overlaps, infer underlying sequence
- Main challenge:
  - (Near-)duplicate sequences

# Finding transcripts (“RNASeq”)

- Get *reads* from cDNA copies of the processed (spliced + edited) transcripts
- *Assemble* to infer transcript sequence
- *Align* to genome sequence
- Main challenges:
  - A given transcript may not be present in all cells
  - Transcripts may be processed in more than one way (isoforms)
  - A transcript may be non-functional!

# Finding sites

- Direct detection of binding events (e.g. ChIPSeq)
  - *but* binding may be non-functional!
- Computational search for clusters of recurring motifs
  - *but* motifs occur frequently by chance, in any large genome!
- Both methods are error-prone, and neither illuminates site *function*. So we also ...



# *Compare* genomes of ...

- a lab organism & a singly mutated variant with an altered phenotype
  - the mutation must then alter (or create!) a site, and
  - the phenotypic change illuminates its function
  - but remember that cells with identical genomes can sometimes have different phenotypes!
    - Tissues in multicellular organisms
- members of a natural population
  - Usually *multiple* genomic and phenotypic differences
  - find correlations (of *recurring* differences) to identify sites that affect a particular phenotype.

- different species
  - *Many* differences
  - *atypically similar* (= “**conserved**”) regions likely represent site clusters in which mutations have been selected against (“purifying selection”)
  - but generally can’t conclude anything about function
    - too many phenotype differences
  - also, many sites may have been *lost*, and *created*, in each lineage

# Major computational tasks

- Comparing & aligning sequences
  - Reads to reads
    - assembly
  - Reads to genomes
    - variant detection
  - Transcripts to genomes
  - Genomes (or portions thereof) to genomes

Appropriate alignment method depends on how similar the sequences are!

- Developing probability models of
  - Genome sequences (sites, and “background”)
  - Sequence evolution
  - Other types of ‘linear’ data associated to the genome (e.g. read depth)and using them to find genomic features.

# This course (approximately!):

- *Suffix arrays* for finding exact matches
- *Background sequence models*
- *Site models*, weight matrices & sequence logos
- Highest weight paths on weighted directed acyclic graphs: *dynamic programming algorithm*
- Finding regions of biased composition (“HMMs lite”)
- Edit graphs & *gapped-alignment algorithms*
- *Hidden Markov models* and applications
  - Parsing genomes (into sites & non-sites)
  - Finding conserved regions

- We do *not* cover (but see Genome 541!)
  - motif-finding methods
  - sequence evolution models
  - more complex machine learning models (e.g. deep neural nets)