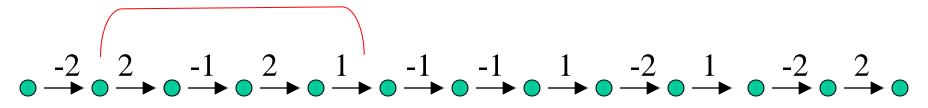
# Lecture 7: Weighted linked lists

- Applications (via *sequence graphs*):
  - regions of atypical residue composition
  - motif clusters
  - read count data
- Finding *multiple* high-scoring paths
- "D-segments"
- Statistical significance

### Weighted Linked Lists (WLLs)

- *WLL* is linked list with weights on each edge simplest kind of WDAG.
- Paths = 'segments' or 'regions'

highest-scoring segment



• Find highest-scoring segments by dynamic programming

– Much better than "brute force" algorithm!

- Beginning & end of best path determine path uniquely, so
  - traceback is unnecessary
  - single pass through list suffices to find best path.

from lecture 6 :

• To reconstruct best path, need "traceback" pointer to immediate predecessor of *v* in best path:

$$T(v) = \begin{cases} v & w(v) = 0\\ \arg \max_{u \in \text{parents}(v)} (w(u) + w((u,v))) & w(v) \neq 0 \end{cases}$$

- in preceding graph, T(v) is the *parent* on *red edge* coming into *v* 
  - if more than one such edge, pick one at random;
  - if no such edge, T(v) = v
- Sometimes useful to record *beginning* of best path:

$$B(v) = \begin{cases} v & w(v) = 0\\ B(T(v)) & w(v) \neq 0 \end{cases}$$

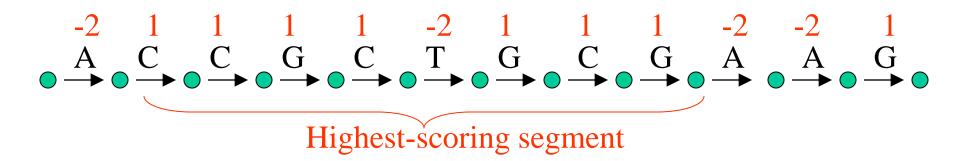
#### **Applications to Sequences**

- A *sequence graph* of a sequence is linked list whose edges are labelled by sequence residues (in order):
- e.g. graph for sequence ACCGCTGCGAAG is:

#### $\overset{A}{\longrightarrow} \overset{C}{\longrightarrow} \overset{C}{\longrightarrow} \overset{G}{\longrightarrow} \overset{C}{\longrightarrow} \overset{C}{\longrightarrow} \overset{T}{\longrightarrow} \overset{G}{\longrightarrow} \overset{C}{\longrightarrow} \overset{G}{\longrightarrow} \overset{G}{\longrightarrow} \overset{G}{\longrightarrow} \overset{A}{\longrightarrow} \overset{A}{\longrightarrow} \overset{G}{\longrightarrow} \overset{G}{\longrightarrow} \overset{G}{\longrightarrow} \overset{C}{\longrightarrow} \overset{G}{\longrightarrow} \overset{C}{\longrightarrow} \overset{G}{\longrightarrow} \overset{C}{\longrightarrow} \overset{G}{\longrightarrow} \overset{G}{\longrightarrow} \overset{C}{\longrightarrow} \overset{G}{\longrightarrow} \overset{G$

### Weighted Sequence Graphs

• If attach weight to each residue, sequence graph becomes a WLL.



• Useful for identifying sequence regions ('target regions') with atypical composition:

- In DNA:
  - -GC-rich regions in AT-rich thermophile genomes
    - generally correspond to RNA genes (Rob Klein & Sean Eddy)
  - -horizontally transferred regions
  - -isochores (mammalian DNA)
- In proteins:
  - -hydrophobic regions (transmembrane segments)
  - hydrophilic regions (loops, intrinsically disordered regions)
  - acidic or basic regions

### 'Optimal' scores

- Assume sequence consists of
  - target regions with residue freqs  $t_r$
  - background regions with residue freqs  $b_r$
  - *independence assumption* applies in both
- *Then* 'best' scoring system to detect the target regions uses LLRs:

 $s(r) = \log(t_r / b_r)$ 

• if residue freqs are unknown, can usually estimate iteratively

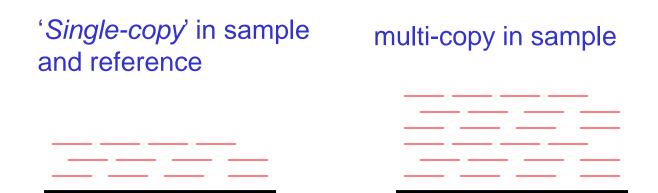
# Can use *non-residue-based* scores to find:

- Regions enriched in particular sequence *motifs*:
  - CpG islands in mammalian genomes
    - positive weight (e.g. +17) to the first C of each CpG, and
    - negative weight (e.g. -1) to every other base
      (This approach was used in *Nature* human genome paper).
  - Regions rich in (known) transcription-factor motifs
  - Optimal scores are LLRs, but now based on 'symbol frequencies' (where symbol = presence/absence of motif)

- Regions targeted by *next-gen read experiments* (symbols = *read counts*)
  - CNVs (Homework 5)
  - Hypersensitive sites
  - CHIP-seq
- Conserved regions in *sequence alignments* (symbols = *alignment columns*)

#### CNVs & Read Depth

- CNV = 'copy number variant'- e.g. region that is single copy in reference sequence but duplicated in sample
- One way to detect: map reads from sample onto reference, look for regions of atypical coverage depth



# HW 5: finding CNVs using D-segments

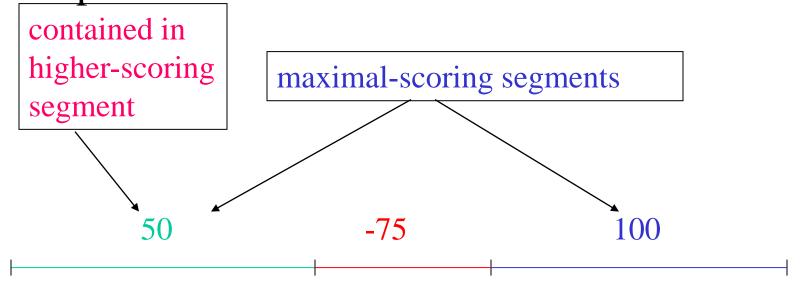
- *data*: next-gen read alignments to genome
- observed symbols: *counts* of # *read starts* at each position  $(0, 1, 2, \ge 3)$ 
  - *frequencies* from **Poisson dist'n** with appropriate mean
- target regions: *heterozygous duplications* 
  - One chrom = reference allele, other is dup
  - Poisson mean = 1.5 X background mean

# Finding *multiple* high-scoring segments

- In general, expect several regions of particular type in a given sequence not just one!
- So want to find multiple high-weight paths in a WDAG
- But not interested in slight perturbations of previously found paths
- One strategy:
  - Find highest-weight path
  - 'Mask it' (remove its edges from graph)
  - Repeat above two steps until scores no longer 'interesting'

- Is there a more efficient algorithm not requiring repeated scans?
  - Ruzzo & Tompa solved for WLLs
  - $-\exists$  solution for arbitrary WDAGs?

- A (*locally-*)*maximal*(*-scoring*) *segment* I is one such that
  - -P1: no subsegment of I has a higher score than I
  - P2: no segment properly containing I satisfies P1
- Example:

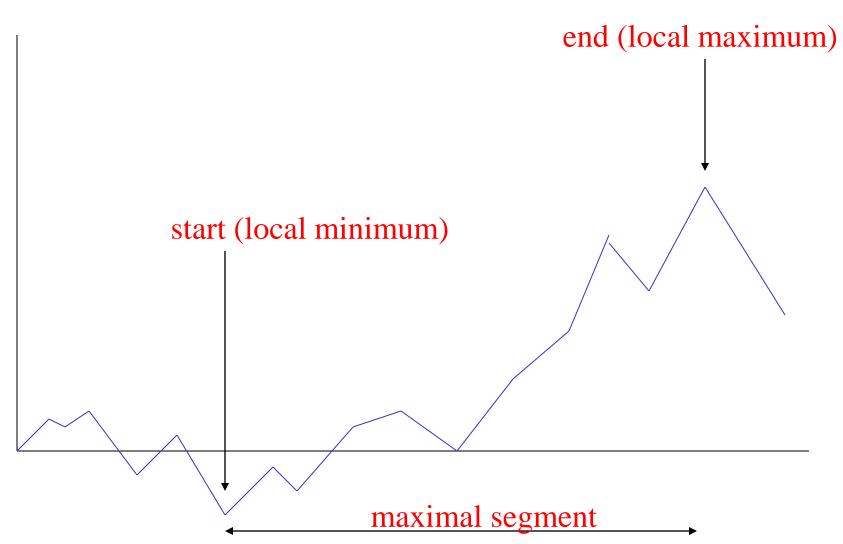


score = 75, but does not satisfy *P1* 

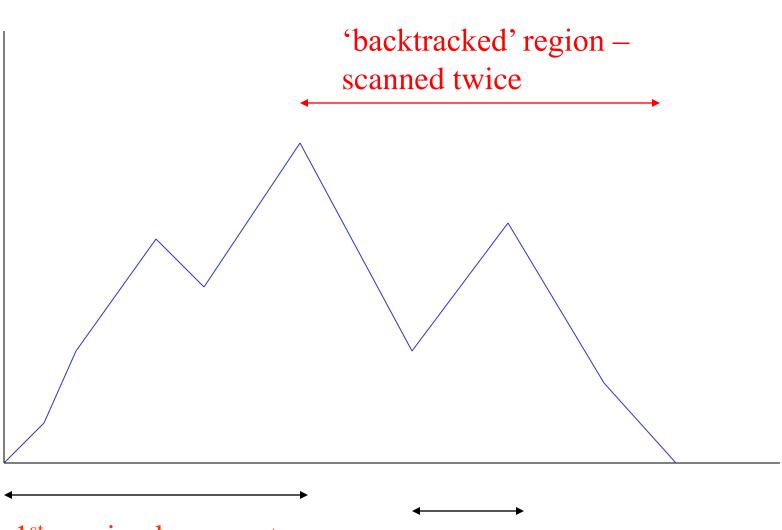
- *Problem*: given S > 0, find all maximal segs of score  $\geq S$
- Segments are *paths* in a linked-list WDAG with *N*+1 vertices and *N* edges
- Highest weight path is found by dynamic programming; in (pseudo-)pseudocode: cumul = max = 0; start = 1; for (i = 1; i ≤ N; i++) { cumul += s[i]; if (cumul ≤ 0) {cumul = 0; start = i + 1;} /\* NOTE RESET TO ZERO \*/ else if (cumul ≥ max) {max = cumul; best\_end = i; best\_start = start;} }

```
if (max \ge S) print best_start, best_end, max
```

# Maximal segments – from cumulative score plot (without 0 resets)



• Can find *all* maximal segs of score  $\geq$  S using following practical (but *non-optimal*) algorithm: cumul = max = 0; start = 1;for  $(i = 1; i \le N; i++)$ cumul += s[i];if (cumul  $\geq$  max)  $\{\max = \operatorname{cumul}; \operatorname{end} = i;\}$ if (cumul  $\leq 0$  or i == N) { if  $(\max \ge S)$ {print start, end, max; i = end; } /\* N.B. MUST BACKTRACK! \*/ max = cumul = 0; start = end = i + 1;



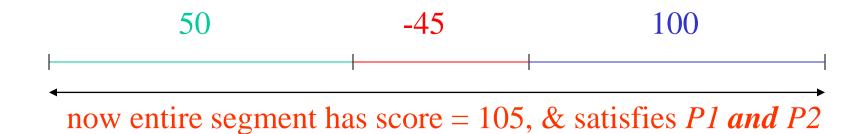
1<sup>st</sup> maximal segment

2<sup>d</sup> maximal segment

- In worst case this is O(N<sup>2</sup>) (because of backtracking),
  - but in practice usually O(N) because a given base is usually traversed only a few times
- Ruzzo-Tompa algorithm *guarantees O(N)*

- undesirable aspect of maximal segments as defined:
  - single maximal seg may contain *two* (or more) highscoring regions, separated by significant negativescoring regions
  - i.e. two possibly biologically distinct target occurrences get merged into one maximal segment

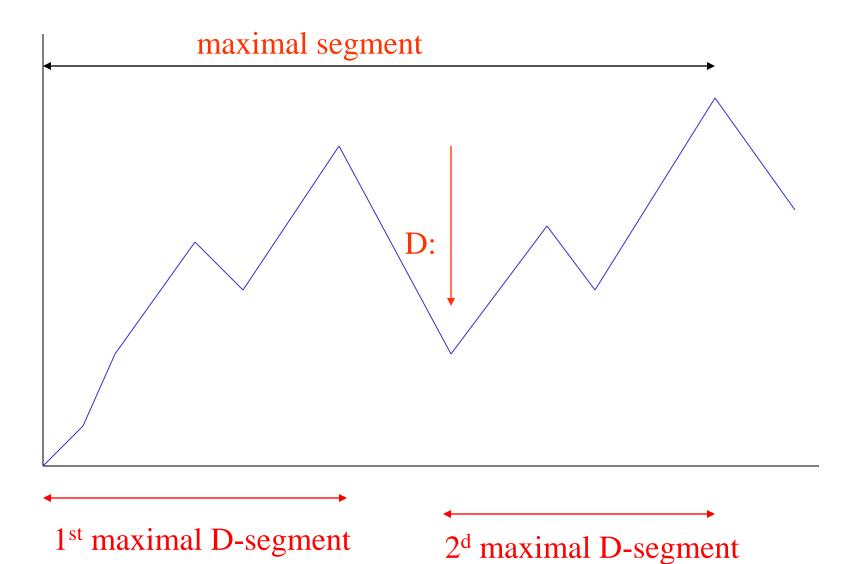
• Example:



#### A better problem!

- to avoid this, have max allowed 'dropoff' D
   < 0</li>
- *D-segment* is segment without any subsegments of score < D</li>
- *maximal D-segment* is D-segment I such that
  - *P1*: no subsegment of I has higher score than I
  - P2: no D-segment properly containing I satisfies P1
- Problem: given  $S (\geq -D)$ , find all maximal D-segs of score  $\geq S$ 
  - (algorithm fails if S < -D)

#### Maximal D-segments



```
O(N) algorithm to find all maximal D-segs:
 cumul = max = 0; start = 1;
 for (i = 1; i \le N; i++)
      cumul += s[i];
      if (cumul \geq max)
           \{\max = \operatorname{cumul}; \operatorname{end} = i;\}
      if (\text{cumul} \le 0 \text{ or cumul} \le \text{max} + D \text{ or } i == N) {
          if (\max \ge S)
             {print start, end, max; }
           max = cumul = 0; start = end = i + 1; /* NO BACKTRACKING
             NEEDED! */
```

- So more biologically relevant problem is also computationally simpler!
- what are appropriate S and D?
  - mainly an empirical question (based on known examples); altho
    - interpretation via 2-state HMM can be useful
    - Karlin-Altschul theory tells when they are 'statistically significant'

### **D-Segments**

- Powerful tool for analyzing 'linear' data
  - Single sequences (incl. motifs, numerical data)
  - Fixed alignment
- Strengths:
  - Very simple to program
  - Very fast, even for mammalian genomes
- Main limitation:
  - Only allows two types of segments ('target' and 'background')
    - Essentially a generalization of 2-state HMMs
    - multi-state HMMs are more flexible

Statistical significance of segment scores

- How often does a given score occur by chance in background sequence?
- Can suggest (but not prove!) biological significance

#### Methods for Assessing Significance of Maximal Segment Scores

- 1. exact prob dist'n
- 2. approximate formula (Karlin-Altschul)
- 3. from simulated sequences
- 4. from real biological 'background' sequences
  - i.e. not having feature in question
- 1, 2, 3 require probability model approximating biological reality; 4 requires an appropriate dataset
- 2 is faster than 1 or 3 (and gives 'intuition'), but involves add'l approximations (ignores 'edge effects')
- 1 requires more complex algorithm

#### Karlin / Altschul approximation

- for  $s(r) = \log_b(t_r / b_r)$ , expected # segments of score  $\ge S$  in (random) backgd seq of length N  $\approx NK b^{-S}$
- for some constant *K* (not depending on *S*)
- Note that  $b^{-S} = b^{-LLR} = 1 / LR$

so (apart from *K*) this is essentially the observation in lecture 5:

# *(from lecture 5) Average* likelihood ratios

- *average LR* (for sites) ≈ *average spacing* between occurrences of 'site-like' sequences *in background*
- So e.g. for 3' splice sites
  - if the average *LR* is 1000, then one expects 'splice-sitelike' sequences to occur on average once per kb *in background sequence*
  - *N.B.* This says nothing about the frequency of *actual* splice sites! (which could be greater or smaller than 1 per kb), and so doesn't by itself provide the probability that an *apparent* splice site is an *actual* site.